

Study of the Effect of Morigina Oleifera on Chromosome Abnormalities in white Mice

Zahraa Taleb Kazem¹, Aseel Rahim Mardan²

¹ Department of Biology, College of Education, University of Al-Qadisiyah.

² Assist. Prof. Dr., Department of Biology, College of Education, University of Al-Qadisiyah.

E: mail: aseel.alaamiri@qu.edu.iq

Abstract

The present work was designed to detect the genetic effects of the aqueous extract of Moringa oleifera leaves and to show the effectiveness of this extract in inhibiting the targeted genetic mutations by the carboplatin. Three concentrations of the extract were chosen to study the genetic effects. The body these concentrations were given for 35 days to white mice and based on some genetic tests that included chromosomal aberrations in bone marrow cells and after reaching the best dose, for example, from Moringa oleifera 1500 gm/ kg which is the dose that gave the least number of chromosome deviations, the effect of this dose is studied In protecting the cells of the bone marrow from the side effects of the carboplatin drug, the effect of the drug carboplatin was studied at the same time period and at a dose of 90 mg / kg of body weight.

Keywords: Chromosome abnormalities, Moringa oleifera, White mice.

1. Introduction

Because of the tremendous cognitive and technical progress in the manufacture of medicines and chemical preparations, modern medicine has proven its effectiveness in treating most of the known diseases, but it soon stopped helpless in the face of some dangerous diseases, which prompted many researchers and scientists to find ways such as herbal medicine to eliminate chronic diseases that did not He is able to find a treatment for it, as its effects have become a civilization of the human race because it may target its genetic material, causing a mutation that can be passed on to generations (Abdulla and Gruber, 2000; Lucock, 1999; Filiberti et al., 1997).

The protective effect of the medicinal plant is mainly due to the fact that it contains a group of chemical compounds that are available in sufficient quantities to limit the development and growth of cancer cells, which gives them the ability to prevent cancer. Due to the recent increasing confrontation towards medicinal plants for preventive or curative use, the Moringa tree, which is characterized by high stability towards self-oxidation, as well as its antioxidant role and inhibitory activity for many types of bacteria and fungi (Alhusnan and Alkahtani, 2019), was chosen. The effectiveness of its leaves lies in the fact that it contains flavonoids, corticosteroids and saponins (Nazmy et al., 2016). Its leaves are also edible and are of high nutritional and therapeutic value due to their rich content of vitamin A, vitamin E, vitamin C and minerals, especially calcium and potassium, and contain many antioxidants, amino acids and carotenoids (Nihad et al., 2019). Moringa leaves are used as an effective treatment for malnutrition because they contain a group of chemicals in the leaves, pods and seeds (Jwa., 2019). Anti-cancer drugs may cause many effects, including the effect on DNA, and this effect results in DNA damage that leads to chromosomal aberrations that cause

chromosomal instability and mutations (Degrassi et al., 2004; Attia, 2008). Carboplatin is one of the most effective drugs in the treatment of cancerous diseases since 1981 (Vijayalaxmi and D'souza, 2004). This drug is used to treat breast cancer (Brambilla et al., 1993), lung cancer and urinary bladder cancer (Hartmann and Lipp, 2003). Among the side effects caused by chemotherapy drugs, including platinum drugs, are hematological toxicity, which affects the bone marrow functions and the production of blood cells, which causes anemia due to a lack of blood cells and platelets (Kamimura et al., 2016).

The current study was proposed and completed to shed light on the genetic effects of carboplatin and the moringa plant, through the use of the aqueous extract of Moringa leaves and its content of effective compounds to reduce and reduce the occurrence of genetic mutations of carboplatin using the CA test for chromosomal aberrations.

Plant collection and identification Moringa leaves

were obtained from the farm of the assistant professor, Dr. Abbas, one of the teachers of the Department of Life Sciences, University of Al-Qadisiyah in October 2021. He was diagnosed by the assistant professor, the doctor, who specializes in plant classification, Dr. Suhaila Hussein Al-Lami, and the plant leaves were cleaned well, then grinded. In the herb grinder to get a fine powder and keep in clean plastic containers until use.

Aqueous extraction method Moringa plant

Fresh Moringa leaves were air dried for 7 days at 30°C and then the leaves were crushed in an electric grinder until a fine powder was obtained. The room after covering it, and then filtering the cell using several layers of medical gauze to get rid of plankton and then placing the solution in clean and sterile metal dishes and drying the extract using the oven for four days at a temperature of 40 degrees Celsius, which gives a greenish-brown color, the weight of

the dried extract was about 22 Then, the dried extract was weighed and reformulated in water pH 6.8 to obtain a stockpile of 200 mg.

Doses and concentrations of aqueous extract Moringa leaves

The plant extract was prepared according to and appropriate to the cytogenetic study in the bone marrow cells of male albino mice. Three concentrations of the aqueous extract were used 1500, 500, 250 mg/kg of body weight, then the mice were administered orally using a syringe Modified for this purpose and preparation The main concentrations The aqueous extract was dissolved in distilled water (Olufunsho et al., 2012).

Carboplatin dose

The carboplatin dose of 90 mg/kg was selected on the basis of body weight and equivalent to the therapeutic human dose of 400 mg/kg (Basit et al., 2021). Carboplatin was dissolved in distilled water and injected into the peritoneal cavity of mice as a single 0.1 ml dose.

Laboratory animals

were used in this study 60 male white mice, their weights ranged between (30-25) grams and their ages ranged between (7-14) days. They were obtained from the animal house of the Department of Life Sciences, College of Science, University of Kufa. The animals were bred in The animal house belongs to the College of Science, University of Al-Qadisiyah. The animals were left for two weeks to acclimatize under laboratory conditions of suitable ventilation and at a temperature of (25) Celsius. That's what animals used in experiments. Design of experimental study Choosing a dose of *Moringa oleifera* extract

The experiment assigns (30) males to six mice per group. This stage included studying the genetic effects of the Moringa extract using three doses according to (Olufunsho et al., 2012) and the dose of carboplatin according to (Basit et al., 2021). The first group, negative control: the animals were left without treatment other than distilled water for 35 days. The second group, the wave control: the rats were injected daily with the carboplatin mutagen in the peritoneal cavity At a concentration of 90 mg kg for 35 days. The third group: Orally administered using a syringe liberated for this purpose, a plant extract Moringa at a concentration of 250 mg kg for 35 days. Fourth group: Orally administered using a syringe modified for this purpose plant extract Moringa at a concentration of 500 mg/kg for 35 days. Fifth group: Orally administered using a syringe Suhoor for this purpose plant extract Moringa at a concentration of 1500 mg kg for 35 days. After the end of the period, the animals were explained for the purpose of conducting genetic tests, and in light of the results of these groups, the optimum dose of the extract was chosen for the purpose of conducting an interaction with one of the genetically mutagenic drugs (Mutagenic) and this drug is Carbopltin, and

the body weight of the experimental animals was calculated before and after treatment with the extract for mice.

The study of the interaction between the optimal dose (mg/kg) of the aqueous extract of Moringa and the drug carboplatin to study the mechanism by which the extract of Moringa works. 6) mice for each group, i.e. a total of (30) proximal mice:

The first group, negative control: the animals were left without treatment except for distilled water for 35 days.

The second group, the wave control: the rats are injected daily with the carbo-platinum airport in the peritoneal cavity at a concentration of 90 mg kg for 35 days.

Group III: Treatment with the plant extract before the mutator Al-Omran dosed the plant extract in the optimal concentration three weeks before the mutator for 35 days of treatment.

Fourth group: Treatment with the plant extract with the mutagenic periodically, the plant extract is dosed at the optimum concentration, and then it is injected with the mutagenic for a period of 35 days from the treatment.

Fifth group - treatment with the plant extract after the muzzar Perch for genetic testing.

Cytogenetic study

Each rat was injected (0.25 ml) of collagen solution at a concentration (0.5 mg/ml) intraperitoneally two hours before killing the animals to stop the animals. M cells at the metaphase.

2- Sacrifice the animals by anesthetizing them with chloroform.

3 - Stabilizing the animals on their ventral side on the autopsy board and the ventral side of the animals, and the thigh area was wiped with 70% ethanol, the animal was dissected by cutting the skin directly and the organs were extracted from their sites

4- Then the femur is taken after cleaning it from the tissues and muscles and it is cut from the middle in a position perpendicular to the edge of the test tube. Using a sterile syringe, 5 ml of phosphate buffer solution (PBS) is injected to wash it and drop the bone marrow into the test tubes.

5- The tubes were centrifuged at a speed of (2000 rpm) for 10 minutes.

6- The supernatant was removed and (5 ml) of potassium chloride (KCl) was added to the precipitate and hypotonic at a concentration 0.075 ml tubes were then incubated in a shaking water bath at (37°C) for (30 minutes).

7- The tubes were extracted from the water bath and built in a centrifuge at a speed of 2000 cycles/minutes) for (10 minutes).

8- The cap was removed, and the prepared fixative solution was gradually added to the precipitate in the form of drops that spilled onto the inner wall of the tube with continuous shaking, then completed the volume of the added stabilizer to reach (5 ml).

9- The tubes were placed at a temperature (4 degrees Celsius) for half an hour for the purpose of fixing the cells

10- The tubes were centrifuged at a speed (2000 rpm) for 5 minutes, then the supernatant was removed, and the cells were suspended again in an appropriate volume (1-1 ml) of cold fixative.

11- The tubes containing the fixed cells were shaken and (8-6) drops of the contents of the tube were dropped onto a clean glass slide vertically from a distance of about (3 feet) to allow the cells to spread well, then the slides were dried on a hot plate (50 m).

Chromosomal aberration test

The slides were examined under an ordinary light microscope under the xenical lens (100). Cells are in the tropics of cell division, when chromosomal aberrations are evident, and can Estimation of the percentage of these aberrations (Allen et al., 1977).

2. Statistical Analysis

The results of the data were analyzed statistically by using the statistical analysis program (SPSS 23) discovery version 2015 by finding the mean +

standard error (meanom), and using the SE M Least significant difference (LSD) test. Statistical data between experimental groups on animals was analyzed using the method one way ANOVA and - two sample T-Test for analysis of control groups). The results are considered significant if the p-value is less than 0.05 (Sorlic, 1995).

3. The Result

The table showed a moral rise of 0.05 >p in the percentage of chromosomal deviations in the positive control treatment of 2.492 compared to negative control treatment of 1.689, The water extract coefficient of the Moringa Oleifera leaves showed a moral decrease of 0.05 >p in the percentage of chromosomal deviations of the three tracks (1500, 500, 250) mg/kg, at 1,175, 1,405, 1,577 respectively compared to negative control, There is also a moral decrease of 0.05 >p in all three transactions of the water extract of Moringa Oleifera leaves compared to positive control

Table (1): Effect of carboplatin and various compositions of the water extract of Moringa Oleifera leaves on the appearance of chromosomal deviations.

Parameters Groups	Deletion % (mean±SD)	Dicentric % (mean±SD)	Acentric % (mean±SD)	Ring % (mean±SD)	Chromosome break% (mean±SD)	Total % (mean±SD)
Control	A 0.2146+0.0737	A 0.5083+0.098	A 0.481+0.1074	A 0.1856+0.0646	A 0.2998+0.1279	A 1.689+0.329
Carboplatin	AB 0.4036+0.1087	A 0.6533+0.0966	A 0.6041+0.1034	A 0.3771+0.0857	AB 0.4541+0.1161	B 2.492+0.297
Moringa 250mg	B 0.2847+0.1172	B 0.3062+0.1151	B 0.3092+0.0736	AB 0.3379+0.1293	AB 0.3392+0.0957	BC 1.577+0.34
Moringa 500mg	B 0.2658+0.1206	B 0.2977+0.1129	B 0.2473+0.1257	AB 0.2986+0.064	AB 0.2959+0.1104	BC 1.405+0.259
Moringa 1500mg	B 0.2441+0.0675	B 0.2206+0.1389	B 0.2152+0.0742	B 0.2484+0.129	B 0.2467+0.1457	C 1.175+0.401
Sign.	Sign.	Sign.	Sign.	Sign.	Sign.	Sign.
P-value	0.046	0.0007	0.00005	0.049	0.0317	0.00002

The table shows a moral rise of 0.05 >p in the proportion of chromosomal deviations in the positive control treatment of 2.5357 compared to negative control of 1.6418, While the rate of deviations in interference coefficients decreased (before, together, after) and the moral decrease was 0.05 >p in interference coefficients, the interference

treatment together was 1.652 and more than two interference factors (before, after)1.941, 2.036 respectively compared to positive control, While there are no moral differences between the interference treatment together with the negative control treatment.

Table (2): effect of interference between carboplatin and water extract of Moringa Oleifera leaves (before, together, after) on the percentage of chromosomal deviations

Parameters Groups	Deletion % (mean±SD)	Dicentric % (mean±SD)	Acentric % (mean±SD)	Ring % (mean±SD)	Chromosome break% (mean±SD)	Total % (mean±SD)
Control	A0.2762+0.131	A0.4818+0.1065	A0.3066+0.1112	A0.2722+0.1188	A0.305+0.1102	A1.6418+0.0562
Carboplatin	A0.3658+0.0793	B0.6392+0.1206	AB0.6102+0.1494	A0.4034+0.1697	B0.5172+0.0655	B2.5357+0.2035
Moringa 1500mg+Car	A0.331+0.0873	B0.4412+0.1349	BC0.4742+0.1539	A0.4076+0.078	B0.3824+0.117	BC2.036+0.256
Car+ Moringa 1500mg	A0.2946+0.1323	B0.4615+0.0804	BC0.403+0.0757	A0.4077+0.0948	B0.3741+0.0878	C1.941+0.348
Carb. = Moringa 1500mg	A0.2577+0.0763	B0.3441+0.1037	C0.3389+0.0786	A0.3408+0.0893	B0.3704+0.0965	C1.652+0.296
Sign.	Non-Sign.	Sign.	Sign.	Non-Sign.	Sign.	Sign.
P-value	0.501	0.008	0.005	0.285	0.035	0.0005

4. Discussion

Studied cellular criteria to choose the optimal dose of the aqueous extract of *Moringa oleifera* leaves chromosomal deviations in chromosomal Aberration in bone marrow (CA) led to various doses of hydroenotic extract *Moringa Oleifera* leaves

To reduce the frequency of chromosomal deviations that appear in the table (1-1). The water extract of *Moringa Oleifera* leaves contains natural compounds that do not have genetic toxicity to bone marrow cells such as flavonoids, soapin, etc. that have led to a decrease in the average number of cells with fine nuclei or chromosomal deviations. Studies have shown that flavonoid compounds significantly reduce total chromosomal deviation and these substances can increase the programmed death of abnormal cells. Since most cells suffer from deviation.

Flavonoid compounds found in plants are of great importance in preventing carcinogenicity by protecting DNA damage and stimulating repair pathways, and regulates many important cellular events such as cell cycle and apoptosis (Twari and Mishra, 2017). Flavonoids are powerful antioxidants and have a wide range of biochemical functions, participating in immune functions and gene expression, capillary and cerebral blood flow and liver function, enzyme activity and platelet aggregation and cholesterol. The beneficial health effects associated with these compounds are believed to reduce the risk of coronary heart disease and various types of cancer mainly from antioxidant activity, including the removal of heavy metal from metal ions and the inhibition of fat peroxide (Formica and Regelson, 1995). Studies conducted by Hedges and Lister, 2007, show that kerosene compounds reduce the onset of cancerous tumors, promote stomach ulcer healing and prevent the proliferation of ovarian, breast and colon cancer cells. Studies have shown that terpenes are effective against various types of cancers, including and colon cancer. Carboplatin cellular tests of carboplatin effect on chromosomal deviations

consider bone marrow to be a rich source of both hematopoietic stem cells and stem cells as well as where adult cell group differentiates (Acton, 2013). These cells are highly sensitive to mutation-causing agents, and are therefore highly sensitive to DNA damage that can cause mutations and genetic disorders and can be particularly dangerous in undifferentiated cells in the bone marrow (Antunes et al., 1999). If these cells survive and multiply, the risk of secondary cancer becomes much higher (Travis et al., 1996). The main objective during chemotherapy is to avoid the destruction of non-cancer cells from the unfavorable side effects of therapeutic drugs. The benefit of using natural edible antioxidants can reduce the incidence of these harmful effects and some may help prevent dna damage caused by these drugs, according to several studies (Mora et al., 2002). The ability of free

radicals to interact in chemically stable molecules with anti-cancer drugs can be used to reduce oxidative stress and thus help prevent oxidative damage to DNA and other cellular molecules (Trem and Smejck et al., 2016). Widespread use of chemotherapy drugs can cause cancer treatment to improve the lives of cancer patients, but some anti-cancer drugs can cause a number of direct symptoms and toxicity, and many anti-cancer drugs can be mutagenic or carcinogenic.

Carboplatin is an anti-tumor drug with side effects. A 2017 Chen et al. study showed that carboplatin caused serious bone marrow damage and the disappearance of bone barriers and hemoid fibrosis. The toxic effect of carboplatin is myelosuppression bone marrow inhibition and the main symptoms of carboplatin bone marrow depression include thromboblating, neutrophils, neutrophils and anaemia, which occurs in 20-40% of people treated with carboplatin and more than 90% in patients treated with a high dose of carboplatin (Groopman and Itri, 1999). Bone marrow suppression depends on the concentration of dose when taking carboplatin because platinum-based drugs cause damage to bone marrow cell DNA (Chen et al., 2017). Carboplatin myeloid toxicity is formed by generating free radicals that attack DNA in bone marrow cells (Moon et al., 2011; Lin et al., 2010). Carboplatin can also cause damage to DNA despite the age of short bone marrow cells (Patra et al., 2018).

The mechanism of action of platinum drugs is linked to DNA cross-linking to toxicity events in hematopoietic stem cells (Weijl et al., 2004). Carboplatin also targets salafi cells in the bone marrow that include part of the hematopoietic stem mesinoid cells (Das et al., 2008). Carboplatin also reduced egg blood cells (Kohono, 1995). Carboplatin has a similar effect to sysplatin but has fewer side effects (Hatfield et al., 2016). Treatment with cisplatin increases SOD activity in bone marrow cells while reducing GSPX level and these changes are associated with low GST, GSH and moral decline in hematopoietic cells and the growth of mesinkin stem cell colonies (Das et al., 2008). Carboplatin also reduces the level of glutathione in the bone marrow (Abd-Allh et al., 2005).

choosing the optimal dose for aqueous extract of *Moringa Oleifera* leaves for the purpose of electing the appropriate concentration of aqueous extract for moringa leaf and using it in the work of the intervention experiment with the drug reviewed the results Statistics in table (1.1) on the effects of different concentrations of the water extract of moringa leaves (1500,500,250 mg/kg) on the genetic criteria studied (cell division coefficient), Chromosomal deviations, micronuclearities, sperm deformities, the 1,500mg/kg dose of moringa leaf aqueous extract was adopted as an optimal dose for interfering trials with the desolate drug carboplatin in mice for the purpose of reducing the effect of chemotherapy on the healthy cells of the animal,

although there are some statistical differences in the results between the treatment of this concentration (1500 mg/kg) and the treatment of negative control. Studied cellular parameters after choosing the optimal doses of aqueous extract of *Moringa Oleifera* leaves

chromosomal deviations Chromosomal aberration results indicated that the percentage of chromosomal deviations led to a gradual decrease in interference coefficients (prior, Together, yet, they make a moral difference from the treatment of positive control, in which the rate of deviations was high. As a result, the water extract of *Moringa Oleifera* leaves in all interference coefficients showed a high and clear efficiency in inhibiting the toxic effects of carboplatin, which reduced the values of chromosomal deviations of bone marrow cells of mice. The treatment of interference together was lower in the proportion of chromosomal deviations and this indicates the efficiency of this treatment in preparing cells to counteract the toxic effects of carboplatin, the inhibitive effectiveness of the water extract of *Moringa Oleifera* leaves in transactions (before, Together) of giving the drug was higher than the effectiveness of inhibition after giving the drug due to the fact that inhibitors that are effective (before, together).

The results of the current study showed that the water extract of *Moringa Oleifera* leaves reduces the genetic toxicity caused by carboplatin. All doses of extract selected from the plant have protected the genetic material in the mother cells of the bone marrow by reducing the percentage of chromosomal deviations, as these cells are highly sensitive to drugs that affect cellular division or interfere with the DNA replication mechanism, as these cells have a high split rate, are responsible for the formation of all blood cells and have a short period of time to divide up to 24 hours, All this qualifies them to be the most affected cells with the chemical treatments used in the treatment of many cancers and to damage this type of cell leads to a sharp decrease in all blood cells and this is one of the important side effects of people treated with chemical properties so the use of such extracts may improve the treatment of tumors by chemical treatments. Furthermore, the study showed that patients receiving anti-tumor agents suffer from bone marrow toxicity that causes cell deficiency in the marrow and reduced the effectiveness of hematopoietic tissue (Barreto et al., 2014). The results indicate the cellular toxicity of carboplatin may be caused by free radicals that attack DNA, resulting in the generation of SOD, GST in marrow cells as treatment with carboplatin has led to a high rate of deviations, such as stretching, breaks and gaps in the bone marrow, and the results show that *Moringa Oleifera* plays a protective role against carboplatin mutations. I suggest using plant antioxidants to reduce chromosomal deviations caused by carboplatin. The results of this study are consistent with (Rao et al., 2001). Who found that the plant-based anti-components of *Moringa Oleifera*

reduced the genetic and cellular toxicity that occurs in mice and rats through radiation exposure, as pre-dare with *Moringa Oleifera* extract and then radiation exposure significantly reduced the percentage of chromosomal deviation and micronuclearity.

Consistent with a (Farag et al., 2018) study that suggested that ethanol extract from *Moringa Oleifera* leaves has therapeutic and protective factors that reduce the toxicity of aflatoxin B1 in mice. It is also consistent with a study (Aboelhassan et al., 2018) who referred to the high protective role of moringa leaf extract against genetic changes resulting from aflatoxin B1 and tissue changes in the liver and kidneys in mice, (Radwan et al., 2015), also emphasized the preventive and therapeutic role of *Moringa Oleifera* leaf extract, reducing genetic and cellular toxicity in mice through treatment with CCl₄, which was used to induce liver damage similar to that caused by hepatitis Fyros. in human patients. The results of our study are consistent with the study showing that low non-toxic doses of *Moringa Oleifera* leaf extract protect bone marrow chromosomes and increase the survival of mice exposed to gamma irradiation on the body, due to the antioxidant vitamin C moringa contained, which is responsible for its antioxidant properties and radiation protection of the extract (Anoop and Rao, 2001). Vegetables and fruits are a good source of antioxidants because they contain many phytochemicals, as these natural factors are supposed to have the potential to reduce cellular toxicity and reduce physiological side effects (Landis-Piwowar and Lyer, 2014; Siddiqui et al., 2006). Vegetables contain phenols, carotenoids and vitamin C (Fimognari et al., 2002). Important biological activities of these substances, such as antioxidant, inflammatory, anti-allergic and tumor activity, have been identified

Flavonoids act as powerful anti-mutation antioxidants that protect healthy cells with rapid division coefficients and free radical abrasions (Samejima et al., 1995). Many fruits and vegetables also prevent chromosomal and DNA damage in animals (Nersesyan and Miyata et al., 2004) It has been clarified by explaining the role of anti-mutation plants, one of which may be related to the containment of many powerful antioxidants by plant products, and the observed protective effect on *Moringa Oleifera* against the cellular toxicity of carboplatin exhaled drug may be the result of the positive role of the water extract of *Moringa Oleifera* leaves and its effective substances such as vitamins, beta-carotene and minerals. Selenium, zinc and other compounds (Galuppo et al., 2014; Azad et al., 2017) that has contributed to reducing chromosomal deviations.

Many components of *Moringa Oleifera*, such as polyphenols and various carotenoids, improve the immune system, induce free radicals and reduce the production of DNA mutations in different mammary cells that have previously been exposed to a variety

of oxidized conditions (Nicolle et al., 2003; Van Breda et al., 2005; Srinivasan et al., 2007; Devaraj et al., 2006). Furthermore, polyphenols found in *Moringa Oleifera* have been presented in other studies to inhibit a particular protein found in the bone marrow and increase antioxidants in sperm (Abdou et al., 2012a). Many micro-components of *Moringa Oleifera* have also been considered as an anti-cancer, and have been presented in other studies to reduce the risk of ovarian cancer, lung cancer, prostate cancer in humans and mice (Cramer et al., 2001; Van Breda et al., 2005; Gitenay et al., 2007)

The results of the current study showed that the water extract of *Moringa Oleifera* leaves reduces the genetic toxicity caused by carboplatin. All doses of extract selected from the plant have protected the genetic material in the mother cells of the bone marrow by reducing the percentage of chromosomal deviations, as these cells are highly sensitive to drugs that affect cellular division or interfere with the DNA topoisomerase mechanism, as these cells have a high split rate, are responsible for the formation of all blood cells and have a short period of time to divide up to 24 hours, All this qualifies them to be the most affected cells with the chemical treatments used in the treatment of many cancers and to damage this type of cell leads to a sharp decrease in all blood cells and this is one of the important side effects of people treated with chemical properties so the use of such extracts may improve the treatment of tumors by chemical treatments. Furthermore, this study revealed that the water extract of *Moringa Oleifera* leaves contains therapeutic and protective factors used to reduce the genetic toxicity of carboplatin in mice.

References

- Basit, A., Zafar, M., Liu, X., Javed, A. R., Jalil, Z., & Kifayat, K. (2021). A comprehensive survey of AI-enabled phishing attacks detection techniques. *Telecommunication Systems*, 76(1), 139-154.
- Abdou Bouba, A., Njintang Yanou, N., Foyet, H., Scher, J., Montet, D., & Mbofung, C. M. (2012). Proximate composition, mineral and vitamin content of some wild plants used as spices in Cameroon.
- Azad, M. B., Abou-Setta, A. M., Chauhan, B. F., Rabbani, R., Lys, J., Copstein, L., ... & Zarychanski, R. (2017). Nonnutritive sweeteners and cardiometabolic health: a systematic review and meta-analysis of randomized controlled trials and prospective cohort studies. *Cmaj*, 189(28), E929-E939.
- Formica, J. V., & Regelson, W. (1995). Review of the biology of quercetin and related bioflavonoids. *Food and chemical toxicology*, 33(12), 1061-1080.
- Hedges, L. J., & Lister, C. E. (2007). The nutritional attributes of *Allium* species. *Crop and food research confidential report*, (1814).
- Cox-Georgian, D., Ramadoss, N., Dona, C., & Basu, C. (2019). Therapeutic and medicinal uses of terpenes. In *Medicinal Plants* (pp. 333-359). Springer, Cham.
- Khan, H., Saeedi, M., Nabavi, S. M., Mubarak, M. S., & Bishayee, A. (2019). Glycosides from medicinal plants as potential anticancer agents: Emerging trends towards future drugs. *Current medicinal chemistry*, 26(13), 2389-2406.
- Tiwari, P. (2017). Mishra (2017) 'Role of Flavonoids in DNA Damage and Carcinogenesis Prevention'. *J Carcinog Mutagen*, 8(4).
- Kroemer, G., Petit, P., Zamzami, N., Vayssière, J. L., & Mignotte, B. (1995). The biochemistry of programmed cell death. *The FASEB Journal*, 9(13), 1277-1287.
- Lucock, M. D., Daskalakis, I., Schorah, C. J., Lumb, C. H., Oliver, M., Devitt, H., ... & Levene, M. I. (1999). Folate-homocysteine interrelations: potential new markers of folate status. *Molecular genetics and metabolism*, 67(1), 23-35.
- Barreto, J. N., McCullough, K. B., Ice, L. L., & Smith, J. A. (2014). Antineoplastic agents and the associated myelosuppressive effects: a review. *Journal of pharmacy practice*, 27(5), 440-446.
- Farag, I. M., Roshdy, H. M., Radwan, H. A., Ghaly, I. S., Salah, S. H., Abdel-Rahim, E. A., & Abdalla, A. M. (2018). Ameliorative role of ethanolic extract of *Moringa oleifera* leaf on aflatoxin B1-induced genotoxicity and biochemical alterations in rats. *Journal of The Arab Society for Medical Research*, 13(1), 60.
- Radwan, H. A., Ghaly, I. S., Farag, I. M., & Ezzo, M. (2015). Protective and therapeutic effect of *Moringa oleifera* leaf extract on DNA damage, cytogenetic changes, sperm abnormalities and high level of MDA induced by CCL4 in rats. *RESEARCH JOURNAL OF PHARMACEUTICAL BIOLOGICAL AND CHEMICAL SCIENCES*, 6(1), 1061-1079.
- Acton, J. M. (2013). *Silver Bullet? Asking the Right Questions about Conventional Prompt Global Strike* (p. 18). Washington, DC: Carnegie Endowment for International Peace.
- Filiberti, R., Giacosa, A., & Brignoli, O. (1997). High-risk subjects for vitamin deficiency. *European Journal of Cancer Prevention: The Official Journal of the European Cancer Prevention Organisation (ECP)*, 6, S37-42.
- Antunes, N. L., Small, T. N., George, D., Boulad, F., & Lis, E. (1999). Posterior leukoencephalopathy syndrome may not be reversible. *Pediatric neurology*, 20(3), 241-243.
- Travis, W. D., Lubin, J., Ries, L., & Devesa, S. (1996). United States lung carcinoma incidence trends: declining for most histologic types among males, increasing among females. *Cancer: Interdisciplinary International Journal of the American Cancer Society*, 77(12), 2464-2470.
- Mora, L. B., Buettner, R., Seigne, J., Diaz, J., Ahmad, N., Garcia, R., ... & Jove, R. (2002). Constitutive activation of Stat3 in human prostate tumors and cell lines: direct inhibition of Stat3 signaling induces apoptosis of prostate cancer cells. *Cancer research*, 62(22), 6659-6666.
- Tremblay, J., & Šmejkal, K. (2016). Flavonoids as potent

scavengers of hydroxyl radicals. *Comprehensive reviews in food science and food safety*, 15(4), 720-738.

Chen, X., Wang, J., Fu, Z., Zhu, B., Wang, J., Guan, S., & Hua, Z. (2017). Curcumin activates DNA repair pathway in bone marrow to improve carboplatin-induced myelosuppression. *Scientific reports*, 7(1), 1-11.

Groopman, J. E., & Itri, L. M. (1999). Chemotherapy-induced anemia in adults: incidence and treatment. *Journal of the National Cancer Institute*, 91(19), 1616-1634.

Weijl, N. I., Elsendoorn, T. J., Lentjes, E. G. W. M., Hopman, G. D., Wipkink-Bakker, A., Zwinderman, A. H., ... & Osanto, S. (2004). Supplementation with antioxidant micronutrients and chemotherapy-induced toxicity in cancer patients treated with cisplatin-based chemotherapy: a randomised, double-blind, placebo-controlled study. *European Journal of Cancer*, 40(11), 1713-1723.

Das, B., Antoon, R., Tsuchida, R., Lotfi, S., Morozova, O., Farhat, W., ... & Baruchel, S. (2008). Squalene selectively protects mouse bone marrow progenitors against cisplatin and carboplatin-induced cytotoxicity in vivo without protecting tumor growth. *Neoplasia*, 10(10), 1105-IN4.

Kohno, Y., Egawa, Y., Itoh, S., Nagaoka, S. I., Takahashi, M., & Mukai, K. (1995). Kinetic study of quenching reaction of singlet oxygen and scavenging reaction of free radical by squalene in n-butanol. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 1256(1), 52-56.

Abd-Allah, A. R., Al-Majed, A. A., Al-Yahya, A. A., Fouda, S. I., & Al-Shabana, O. A. (2005). L-Carnitine halts apoptosis and myelosuppression induced by carboplatin in rat bone marrow cell cultures (BMC). *Archives of toxicology*, 79(7), 406-413.

Aboelhassan, D. M., Hafiz, N. A., Darwish, H. R., Shabana, M. E., Eshak, M. G., Hassanane, M. M., ... & Abdalla, A. M. (2018). Enhancing effects of Moringa oleifera leaf extract on carcinogenic Aflatoxin B1-induced genetic alterations, haematoxicity and histological changes in liver and kidney of rats. *BIOSCIENCE RESEARCH*, 15(2), 814-833.

Rao, A. V., Devi, P. U., & Kamath, R. (2001). In vivo radioprotective effect of Moringa oleifera leaves.

Anoop, M. B., Rao, K. B., Rao, T. V. S. R., & Gopalakrishnan, S. (2001). International standards for durability of RC structures: Part 1- A critical review. *Indian Concrete Journal*, 75(9), 559-569.

Landis-Piwowar, K. R., & Iyer, N. R. (2014). Cancer chemoprevention: current state of the art. *Cancer growth and metastasis*, 7, CGM-S11288.

Siddiqui, S. A., Potewar, T. M., Lahoti, R. J., & Srinivasan, K. V. (2006). Ionic liquid promoted facile one-pot synthesis of 1-pyridylimidazo [1, 5-a] pyridines from dipyridylketone and aryl aldehydes. *Synthesis*, 2006(17), 2849-2854.

Fimognari, C., Nüsse, M., Cesari, R., Iori, R., Cantelli-Forti, G., & Hrelia, P. (2002). Growth inhibition, cell-cycle arrest and apoptosis in human T-cell leukemia

by the isothiocyanate sulforaphane. *Carcinogenesis*, 23(4), 581-586.

Chu, X., Liu, A. Z., Papen, G., Gardner, C. S., Kelley, M., Drummond, J., & Fugate, R. (2000). Lidar observations of elevated temperatures in bright chemiluminescent meteor trails during the 1998 Leonid shower. *Geophysical research letters*, 27(13), 1815-1818.

Samejima, K., Kanazawa, K., Ashida, H., & Danno, G. I. (1995). Luteolin: a strong antimutagen against dietary carcinogen, Trp-P-2, in peppermint, sage, and thyme. *Journal of agricultural and food chemistry*, 43(2), 410-414.

Nersesyan, A., Mišák, M., Cherkas, A., Serhiyenko, V., Staudinger, M., Holota, S., ... & Knasmüller, S. (2021). Use of micronucleus experiments for the detection of human cancer risks: a brief overview. *Proceeding of the Shevchenko Scientific Society. Medical Sciences*, 65(2).

Galuppo, R., Maynard, E., Shah, M., Daily, M. F., Chen, C., Spear, B. T., & Gedaly, R. (2014). Synergistic inhibition of HCC and liver cancer stem cell proliferation by targeting RAS/RAF/MAPK and WNT/ β -catenin pathways. *Anticancer research*, 34(4), 1709-1713.

Azad, M. B., Abou-Setta, A. M., Chauhan, B. F., Rabbani, R., Lys, J., Copstein, L., ... & Zarychanski, R. (2017). Nonnutritive sweeteners and cardiometabolic health: a systematic review and meta-analysis of randomized controlled trials and prospective cohort studies. *Cmaj*, 189(28), E929-E939.

Nicolle, C., Cardinault, N., Aprikian, O., Busserolles, J., Grolier, P., Rock, E., ... & Révész, C. (2003). Effect of carrot intake on cholesterol metabolism and on antioxidant status in cholesterol-fed rat. *European Journal of Nutrition*, 42(5), 254-261.

van Breda, E. J., van der Worp, H. B., van Gemert, H. M. A., Algra, A., Kappelle, L. J., van Gijn, J., ... & Dippel, D. W. (2005). PAIS: paracetamol (acetaminophen) in stroke; protocol for a randomized, double blind clinical trial. [ISRCTN 74418480]. *BMC Cardiovascular Disorders*, 5(1), 1-7.

Srinivasan, M., Sudheer, A. R., & Menon, V. P. (2007). Ferulic acid: therapeutic potential through its antioxidant property. *Journal of clinical biochemistry and nutrition*, 40(2), 92-100.

Devaraj, S., Autret, B. C., & Jialal, I. (2006). Reduced-calorie orange juice beverage with plant sterols lowers C-reactive protein concentrations and improves the lipid profile in human volunteers. *The American journal of clinical nutrition*, 84(4), 756-761.

Abdou, M., Hamill, L., & Gilbert, N. (2012). Designing and building an agent-based model. In *Agent-based models of geographical systems* (pp. 141-165). Springer, Dordrecht.

Cramer, W., Bondeau, A., Woodward, F. I., Prentice, I. C., Betts, R. A., Brovkin, V., ... & Young-Molling, C. (2001). Global response of terrestrial ecosystem structure and function to CO₂ and climate change: results from six dynamic global vegetation models. *Global change biology*, 7(4), 357-373.

- van Breda, E. J., van der Worp, H. B., van Gemert, H. M. A., Algra, A., Kappelle, L. J., van Gijn, J., ... & Dippel, D. W. (2005). PAIS: paracetamol (acetaminophen) in stroke; protocol for a randomized, double blind clinical trial.[ISCRN 74418480]. *BMC Cardiovascular Disorders*, 5(1), 1-7.
- Gitenay, D., Lyan, B., Rambeau, M., Mazur, A., & Rock, E. (2007). Comparison of lycopene and tomato effects on biomarkers of oxidative stress in vitamin E deficient rats. *European journal of nutrition*, 46(8), 468-475.
- Khan, S., Basit, A., Hafeez, M. B., Irshad, S., Bashir, S., Bashir, S., ... & Li, Y. (2021). Moringa leaf extract improves biochemical attributes, yield and grain quality of rice (*Oryza sativa* L.) under drought stress. *Plos one*, 16(7), e0254452.
- Awodele, O., Oreagba, I. A., Odoma, S., da Silva, J. A. T., & Osunkalu, V. O. (2012). Toxicological evaluation of the aqueous leaf extract of Moringa oleifera Lam.(Moringaceae). *Journal of ethnopharmacology*, 139(2), 330-336.
- Allen, J. (1977). Short term spectral analysis, synthesis, and modification by discrete Fourier transform. *IEEE Transactions on Acoustics, Speech, and Signal Processing*, 25(3), 235-238.
- Sorlie, P. D., Backlund, E., & Keller, J. B. (1995). US mortality by economic, demographic, and social characteristics: the National Longitudinal Mortality Study. *American Journal of Public Health*, 85(7), 949-956.
- Abdulla, M., & Gruber, P. (2000). Role of diet modification in cancer prevention. *Biofactors*, 12(1-4), 45-51.
- Nihad, K., Berwal, M. K., Hebbar, K. B., Bhat, R., Haris, A. A., & Ramesh, S. V. (2019). Photochemical and biochemical responses of heliconia (*Heliconia stricta* 'Iris') to different light intensities in a humid coastal environment. *Horticulture, Environment, and Biotechnology*, 60(6), 799-808.
- Al-Husnan, L., Al-Kahtani, M., & Farag, R. (2019). Molecular Characterization of Fumonisin Mycotoxin Genes of *Fusarium* sp Isolated from Corn and Rice Grains. *Sultan Qaboos University Journal for Science [SQUJS]*, 24(2), 78-87.
- Degrassi, F., Fiore, M., & Palitti, F. (2004). Chromosomal aberrations and genomic instability induced by topoisomerase-targeted antitumour drugs. *Current Medicinal Chemistry-Anti-Cancer Agents*, 4(4), 317-325.
- Attia, M. A. (2008). *Handling Arabic morphological and syntactic ambiguity within the LFG framework with a view to machine translation*. The University of Manchester (United Kingdom).
- Vijayalaxmi, K. K., & D'souza, M. P. (2004). Studies on the genotoxic effects of anticancer drug carboplatin in vivo mouse. *International Journal of Human Genetics*, 4(4), 249-255.
- Brambilla, E., Gazzeri, S., Moro, D., de Fromental, C. C., Gouyer, V., Jacrot, M., & Brambilla, C. (1993). Immunohistochemical study of p53 in human lung carcinomas. *The American journal of pathology*, 143(1), 199.
- Hartmann, J. T., & Lipp, H. P. (2003). Toxicity of platinum compounds. *Expert opinion on pharmacotherapy*, 4(6), 889-901.
- Kamimura, H. A., Wang, S., Chen, H., Wang, Q., Aurup, C., Acosta, C., ... & Konofagou, E. E. (2016). Focused ultrasound neuromodulation of cortical and subcortical brain structures using 1.9 MHz. *Medical physics*, 43(10), 5730-5735.
- Jwa, H., Oh, D., Park, K., Kang, J. M., & Lim, H. (2019). exbake: Automatic fake news detection model based on bidirectional encoder representations from transformers (bert). *Applied Sciences*, 9(19), 4062.
- Nazmy, S., Hassan, B., Nihad, A. A., Abeer, E., Abd Elhamid, E. M., & Farid, M. (2016). Biochemical studies on Moringa oleifera leaves extract. *Journal of Biology, Agriculture and Healthcare*, 6(16), 33-42.
- Moon, R. J., Martini, A., Nairn, J., Simonsen, J., & Youngblood, J. (2011). Cellulose nanomaterials review: structure, properties and nanocomposites. *Chemical Society Reviews*, 40(7), 3941-3994.
- Lin, P., Engling, G., & Yu, J. Z. (2010). Humic-like substances in fresh emissions of rice straw burning and in ambient aerosols in the Pearl River Delta Region, China. *Atmospheric Chemistry and Physics*, 10(14), 6487-6500.
- Patra, D. K., Pradhan, C., & Patra, H. K. (2018). Chelate based phytoremediation study for attenuation of chromium toxicity stress using lemongrass: *Cymbopogon flexuosus* (nees ex steud.) W. Watson. *International journal of phytoremediation*, 20(13), 1324-1329.
- Landis-Piwowar, K. R., & Iyer, N. R. (2014). Cancer chemoprevention: current state of the art. *Cancer growth and metastasis*, 7, CGM-S11288.
- Olufunsho, A., & Alade, A. (2012). Investigation of lipid peroxidation as probable mechanism of rifampicin toxicity in vivo. *Annals of Neurosciences*, 19(2), 68.